INHIBITION OF EPIDERMAL GROWTH FACTOR-INDUCED ACTIVATION OF PHOSPHOLIPASE C BY PSI-TECTORIGENIN

Sir:

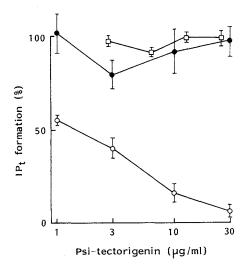
We have isolated psi-tectorigenin from *Nocardiopsis*, as an inhibitor of phosphatidylinositol turnover (PI turnover). It inhibited epidermal growth factor (EGF)-induced inositol incorporation into inositol lipids with an IC $_{50}$ of about 1.0 μ g/ml in human epidermoid carcinoma A431 cells¹⁾. We have studied the mechanism of action and found that psi-tectorigenin acts on the activation of phospholipase C, a rate limiting enzyme of PI turnover.

First, we analyzed the effect of psi-tectorigenin on agonist-induced inositol phosphates production. A431 cells (3×10^5) grown for 16 hours beforehand in 24-well plates were prelabeled with myo- $\lceil^3H\rceil$ inositol (1 μ Ci/ml, 18.3 Ci/mmol, Amersham) for 24 hours in inositol-free Dulbecco's modified EAGLE's medium (DMEM) containing 10% dialyzed calf serum. NIH3T3 cells were grown for 2 days before addition of inositol. Then, the medium was removed and the cells were preincubated for 15 minutes in 0.5 ml of inositol-free DMEM containing 30 mm LiCl. Psi-tectorigenin was then added, and after 15 minutes EGF (400 ng/ml) or 20 μm of ATP was added to A431 cells, or bombesin (10 μm) was added to NIH3T3 cells, and each incubation was continued for a further 5 minutes. The reaction was terminated by the addition of ice-cold 10% HClO₄, and the mixture was then neutralized by addition of 1.53 m KOH in 75 mm HEPES. The solution was kept on ice for 15 minutes after which it was centrifuged at 10,000 rpm for 10 minutes at 4°C. The supernatant was applied to an Amprep SAX column (Amersham), and the column was then washed with water and eluted with 0.17 M KHCO₃. The obtained eluate which contains inositol phosphates (IP+ IP₂+IP₃) was counted in a liquid scintillation counter. Psi-tectorigenin inhibited EGF-induced inositol phosphates production in a dose-dependent manner and the IC₅₀ value was about $1.0 \,\mu\text{g/ml}$, and complete inhibition was observed at 30 µg/ml (Fig. 1). However, psi-tectorigenin did not inhibit ATP-induced formation of inositol phosphates in A431 cells. It also showed no effect on bombesininduced inositol phosphates formation in NIH3T3 cells (Fig. 1). Both ATP and bombesin are considered to act by a tyrosine kinase-independent mechanism^{2,3)}.

Next we measured the effect of psi-tectorigenin on in vitro phospholipase C activity. The assay method was described before⁴⁾. Crude homogenate after removal of nuclear fraction of A431 cells treated with EGF (400 ng/ml) for 10 minutes was used as a phospholipase C preparation. It was incubated with [3H]phosphatidylinositol 4,5-diphosphate ([3H]PIP₂) in the presence or absence of various concentrations of psi-tectorigenin at 37°C for 15 minutes. The reaction was terminated by the addition of 10% TCA and 1% bovine serum albumin. The precipitate was removed by centrifugation, and phospholipase C activity was evaluated by the radioactivity of the TCA soluble fraction containing [3H]IP₃. Psi-tectorigenin did not affect the activity of phospholipase C up to $10 \mu g/ml$. These results indicated that psi-tectorigenin inhibited phospholipase C indirectly.

Then we looked into the effect of psi-tectorigenin on the activation of phospholipase C. We found that the *in vitro* phospholipase C activity in homogenate

Fig. 1. Effects of agonist-induced inositol phosphate production by psi-tectorigenin in A431 cells or NIH3T3 cells.



After pretreatment of cells with verious concentrations of psi-tectorigenin for 15 minutes at 37°C, EGF (400 ng/ml; \odot) or ATP (20 μ M; \odot) was added to A431 cells for 5 minutes. Bombesin (10 μ M; \Box) was added to NIH3T3 cells. Total inositol phosphates were measured as described in text. The 100% and 0% values (dpm) were 1,408.7 \pm 16.4 and 504.4 \pm 101.5, 1,281.48 \pm 45.98 and 889.15 \pm 40.13, and 2,935.45 \pm 323.94 and 1,304.31 \pm 225.38, for EGF, ATP and bombesin, respectively. Values are means \pm SD of triplicate samples.

prepared from EGF-treated A431 cells was 4-fold higher than that from untreated cells, and that EGF failed to activate the enzyme in the presence of psi-tectorigenin (Table 1). Thus, psi-tectorigenin was shown to inhibit the activation step of phospholipase C.

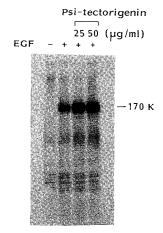
Table 1. Inhibition of EGF-induced phospholipase C (PLC) activation by psi-tectorigenin.

Addition	PLC activity (dpm)
None	332.8 ± 14.2
+ Psi-tectorigenin	547.2 ± 55.0
+EGF	1338.0 ± 14.2
+EGF+psi-tectorigenin	609.6 ± 11.5*

A431 cells were treated or not with psi-tectorigenin (25 μ g/ml) in the presence of Na₃VO₄ (100 μ m) for 15 minutes, and then EGF (400 ng/ml) was added. After 5 minutes, the cells were collected, homogenized and centrifuged at $500 \times g$ for 10 minutes. The phospholipase C activity in the supernatant obtained was determined as described in text. Values are means \pm SD of triplicate samples.

* P<0.02.

Fig. 2. Effect of psi-tectorigenin on tyrosine phosphorylation of EGF receptor.



A431 cells (3×10^5) grown for 16 hours beforehand in 35 mm dish were prelabeled for 4 hours with $100 \,\mu \text{Ci}$ of ^{32}Pi in phosphate-free DMEM containing 4% dialyzed serum. Psi-tectorigenin was added to cells and preincubated for 15 minutes, then EGF ($100 \, \text{ng/ml}$) was added. After 5 minutes, the cells were solubilized with 0.5 ml of cold RIPA buffer⁶. Aliquot of cell lysate was incubated with Sepharose-linked anti-phosphotyrosine monoclonal antibody (Oncogene Science, Inc.) at 4°C overnight. Immune complex obtained was electrophoresed on SDS-PAGE. The tyrosine phosphorylated proteins were detected by autoradiography.

Activation of phospholipase C by EGF is considered to involve EGF receptor tyrosine kinase activation^{3,5)}. In fact, erbstatin, a tyrosine kinase inhibitor, inhibits EGF-induced phospholipase C activation4). However, psi-tectorigenin does not inhibit EGF-induced receptor autophosphorylation as detected by EGF receptor antibody1) and we have confirmed it by using anti-phosphotyrosine antibody (Fig. 2). Psi-tectorigenin at 25 and 50 μg/ml rather increased tyrosine phosphorylation of the receptor slightly. Therefore, psi-tectorigenin should inhibit another process for activation of phospholipase C rather than tyrosine kinase. The mechanism of phospholipase C activation may include a more complex pathway. Psi-tectorigenin may be a suitable tool to elucidate this complex mechanism.

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